

A Standardized System for Grading
Mutations in *Mycobacterium tuberculosis* for
Association with Drug Resistance

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“A lack of user-friendly **data analysis and interpretation tools** has been frequently cited as a major barrier to routine use of WGS techniques.”





“Our experience with WGS in the clinical context of drug-resistant TB highlights **the urgent need for an internationally recognized database for standardized genotype-phenotype correlation interpretation** that can be added to and interrogated by clinicians and microbiologists who will be increasingly accessing WGS platforms at the local hospital level.”

The ReSeqTB Solution: A Standardized System for Grading Mutations

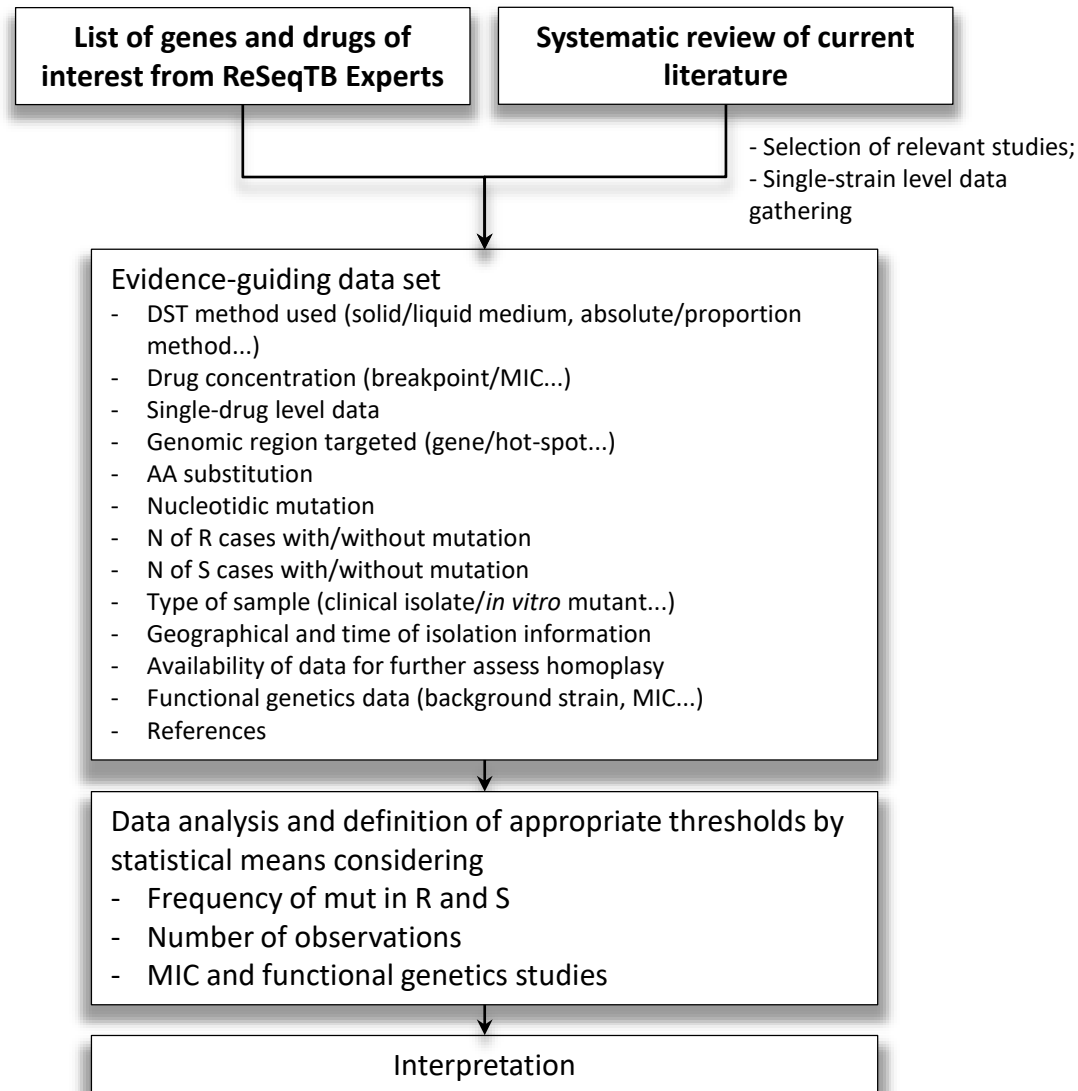
- Driven by Experts and the community
- Grading criteria for confidence binning of mutations
 - Frequency of a mutation in phenotypically resistant or sensitive strain
 - Homoplasmy – similar trait acquired by isolates from different lineages
 - Associated MIC values and the degree of change versus a sensitive strain
 - Functional genetics associated with the mutation
- Every SNP will be associated with a “score” or a “grading index” according to the information available
- Grading drug resistance associated mutations
 - 5 bins based upon the strength of associated data for individual mutations
 - Bins are defined using a statistical approach and then revised according to the expert panels confidence of association with resistance of a mutation

Observed frequency of a mutation found in phenotypically resistant of sensitive strain

- Literature data were used to calculate the frequency of each mutation in resistant (DR) and susceptible (DS) MTBC isolates
- Likelihood ratio (LR) and odd ratio (OR) were used; p-values and 95% confidence intervals associated with LR and OR have been also considered
- Thresholds used most commonly in evidence-based medicine have been adapted to grade *M. tuberculosis* mutations:

LR+ v OR p- value value	Interpretation	Symbol
<0.05 ≥ 10	High Confidence for association with resistance – strong association of the mutation with phenotypic drug resistance; sufficient evidence that the mutation confers or is strongly associated with drug resistance	
<0.05 $5 \leq \dots < 10$	Moderate Confidence for association with resistance – moderate association of the mutation with phenotypic drug resistance; additional data desirable for improved evidence that the mutation confers or is strongly associated with drug resistance	
<0.05 $1 \leq \dots < 5$	Minimal Confidence for association with resistance – weak association of the mutation with phenotypic drug resistance; inconclusive evidence that the mutation confers or is strongly associated with drug resistance. Substantial additional data required	
<0.05 < 1	No association with resistance – No evidence of association between the mutation and drug resistance	
≥ 0.05 -	Indeterminate – no statistically significant threshold reached; additional data required	Indeter

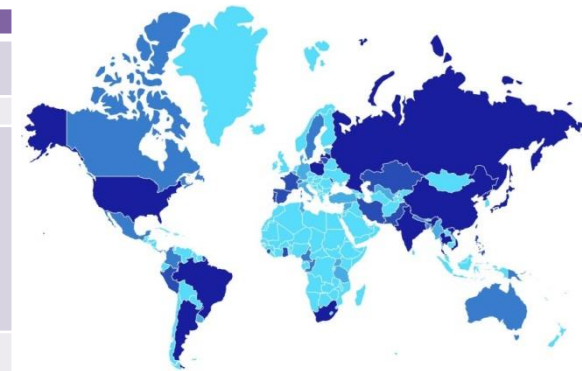
Implementation – Systematic review of literature data



Implementation – Systematic review of literature data

- 11 drugs, 16 genomic loci
- 1748 unique papers screened
- Data: sequencing AND phenotypic DST
- Data from up to 43 countries and up to 13,424 isolates

Drug	Collected data				Number of studies	
	loci of interest	Total # of isolates	Isolation time frame	Represented countries	Screened	Included
Rifampicin (R)	<i>rpoB</i>	13424	1999-2014	37	459	95
Isoniazid (H)	<i>katG</i>	11847	1992-2014	42	650	127
	<i>inhA-mabA</i>	9407				
	<i>furA</i>	361				
	<i>mshA</i>	288				
Ethionamide & prothionamide (ETO/PTO)	<i>inhA-mabA</i>	346	1991-2013	36	243	75
	<i>ethA</i>	181				
	<i>mshA</i>	117				
Ofloxacin (OFX)	<i>gyrA</i>	5911	1990-2014	36	378	81
	<i>gyrB</i>	3078				
Moxifloxacin (MFX)	<i>gyrA</i>	1019	1985-2013	43	423	104
	<i>gyrB</i>	735				
Levofloxacin (LFX)	<i>gyrA</i>	449	1985-2013	43	423	104
	<i>gyrB</i>	218				
Pyrazinamide (Z)	<i>pncA</i>	4949	1990-2014	36	378	81
Streptomycin (S)	<i>rpsL</i>	3263	1985-2013	43	423	104
	<i>tap</i>	0				
	<i>rrs</i>	2598				
	<i>whiB7</i>	0				
	<i>gidB</i>	812				
Amikacin (AM)	<i>rrs</i>	2105	1985-2013	43	423	104
Capreomycin (CM)	<i>rrs</i>	2533				
		<i>tlyA</i>	1854	1985-2013	43	423
Kanamycin (KM)	<i>rrs</i>	1727				
	<i>tap</i>	56				
	<i>eis</i>	2029				
	<i>whiB7</i>	56				



Implementation – Results

Drug (phenotypic testing)		Gene	High confidence mutations	Moderate confidence mutations	Minimal confidence mutations	No association with resistance
First-line	R	<i>rpoB</i>	D516A, D516F, D516G, D516G+L533P, D516ins, D516N, D516V, D626E, Del N518, F505V+D516Y, F514dupl, H526C, H526D, H526F, H526G, H526L, H526R, H526Y, M515I+D516Y, Q513-F514ins, Q513H+L533P, S512T, Q513K, Q513L, Q513P, S522Q, S531F, S531L, S531Q, S531W	D516Y, H526P, L533P, S522L	H526N, I572F, L511P	
	H	<i>inhA-mabA</i>		c-15t		g-47c, t-80g, T4I, g-102a
		<i>katG</i>	S315I, S315N, S315T, Pooled frameshifts and premature Stop codons			A110V, L499M, R463L
		<i>furA</i>				L68F
		<i>mshA</i>			N111S, A187V	
Second-line (group A)	MFX	<i>gyrA</i>	A90V, D94A, D94G, D94N, D94Y, G88C, S91P			E21Q, G247S, G668D, S95T, V712L
	OFX/LFX	<i>gyrA</i>	A90V, D94A, D94G, D94H, D94N, D94Y, G88A, G88C, S91P	D89N		E21Q, G247S, G668D, S95T, T80A, V712L
		<i>gyrB</i>	A504V, E459K			
Second-line (group B)	AM	<i>rrs</i>	a1401g, g1484t			
	KM	<i>eis</i>	c-14t, g-10a		c-12t, g-37t	a1338c
		<i>rrs</i>	a1401g, c1402t, g1484t			
	CM	<i>rrs</i>	a1401g, c1402t, g1484t			c517t
		<i>tlyA</i>	N236K, Pooled frameshifts and premature Stop codons			D149H
	S	<i>rpsL</i>	K43G, K43R, K43T, K88Q, K88R, T40I			
		<i>rrs</i>	a514c, a514t, c462t, c513t, c517t			
<i>gidB</i>					E92D, L16R, V110G, Pooled frameshifts and premature Stop codons	

Implementation – Results

Drug (phenotypic testing)		Gene	High confidence mutations	Moderate confidence mutations	Minimal confidence mutations	No association with resistance
Second-line (group C)	ETO/PTO	<i>inhA</i>	c-15t+I194T, c-15t+S49A	c-15t		
		<i>ethA</i>				Q347Stop
Second-line (group D)	Z	<i>pncA</i>	a-11g, A134V, A3E, A46V, C138Y, C14R, C72R, D12A, D12N, D49G, D49N, D63G, D8E, D8G, D8N, F94L, F94S, G108R, G132A, G132D, G132S, G162D, G17D, G24D, G97C, G97D, G97S, H137P, H51Q, H51R, H57D, H57P, H57R, H57Y, H71D, H71Q, H71Y, H82R, I6T, indel - R148ins (inframe), K96N, K96R, L116P, L116R, L120P, L151S, L159P, L172P, L19P, L4S, L85P, L85R, M175T, M175V, P54S, P62L, P62Q, Q10P, Q141P, R123P, S104R, S59P, S66P, S67P, t-12c, T135N, T135P, T142A, T142K, T142M, T160P, T168P, T76P, t-7c, V125F, V125G, V128G, V139G, V139L, V155G, V180F, V180G, V7G, W68C, W68R, Y103H, Y34D, Pooled frameshifts and premature Stop codons	A171E, K96E, K96T, M175I, P54L, Q10R, W68G	D12G, F58L, H71R, I133T, V139A	I31T, I6L, indel - c-125del, K48T, L35R, T114M, T47A

- 389 mutations (including frameshifts and premature stop codons) had minimal to high confidence values with regards to predicting drug resistance in MTBC

Advantages of the Grading System

Mutations not associated with DR

Drug	Gene	Mutation
	<i>katG</i>	A110V, L499M, R463L
H	<i>inhA-mabA</i>	g-102a, g-47c, t-80g, T4I
	<i>mshA</i>	A187V, N111S
MFX	<i>gyrA</i>	E21Q, G247S, G668D, S95T, V712L
OFX, LFX	<i>gyrA</i>	E21Q, G247S, G668D, S95T, T80A, V712L
KM	<i>rrs</i>	a1338c
	<i>rrs</i>	c517t
CM	<i>tlyA</i>	D149H
	<i>rrs</i>	a1401g
S	<i>gidB</i>	E92D, L16R, V110G, frameshifts and premature Stop codons
Z	<i>pncA</i>	I31T, I6L, c-125del, K48T, L35R, T114M, T47A
ETO/PTO	<i>ethA</i>	Q347Stop

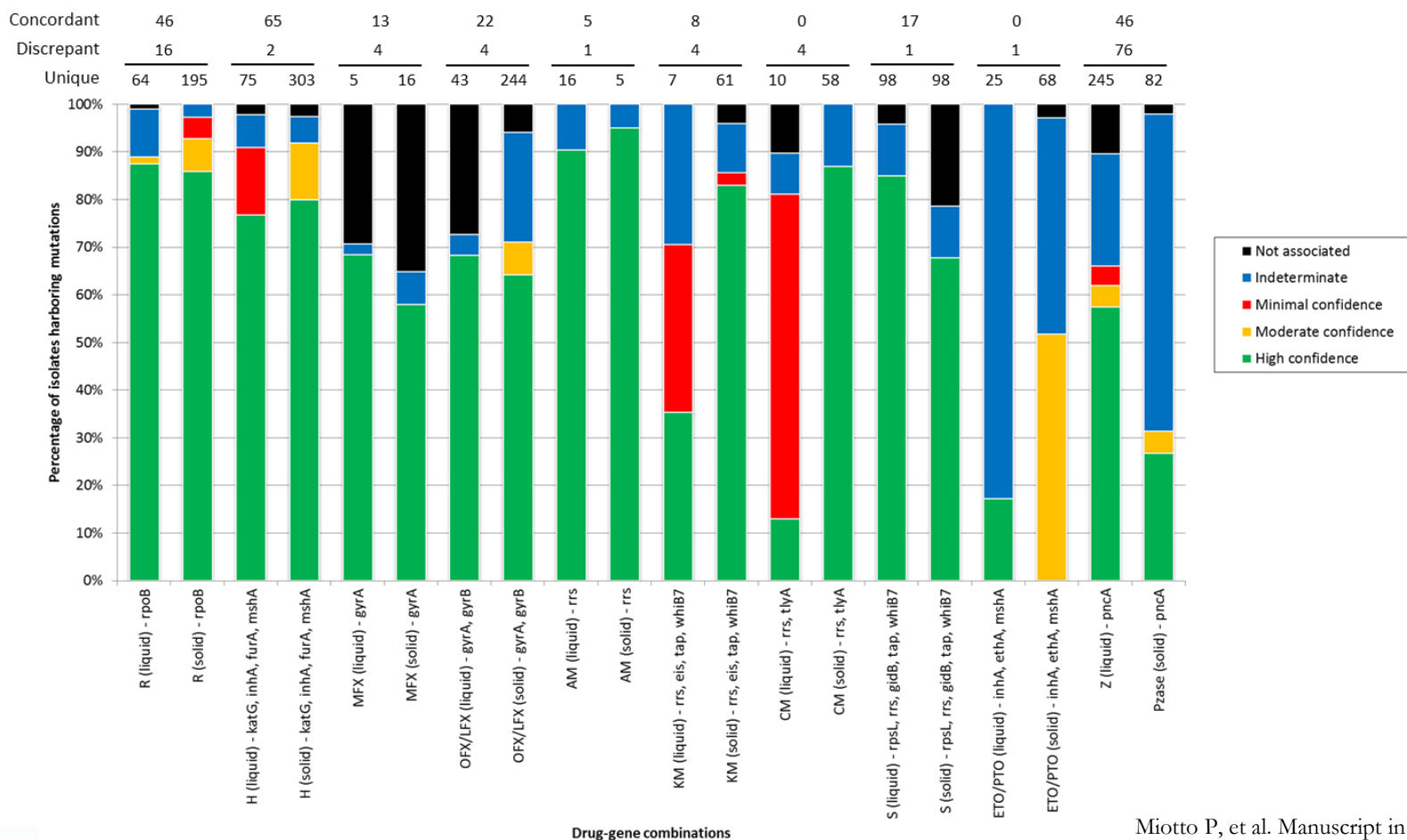
Identification of potential phenotypic testing reproducibility issues

Drug	Gene	Mutation	Found in DR (TP)	Found in DS (FP)	Sensitivity %	Sensitivity 95% CIs	Specificity %	Specificity 95% CIs	Accuracy %	Accuracy 95% CIs	Confidence value
R	<i>rpoB</i>	S531L	4416	20	53.2	52.2-54.3	99.6	99.4-99.8	71.0	70.2-71.7	●
H	<i>katG</i>	S315T	4130	30	53.6	52.5-54.7	96.2	94.6-97.4	57.6	56.5-58.6	●
OFX/OFX	<i>gyrA</i>	A90V	397	4	10.9	9.9-11.9	99.9	99.7-100.0	55.6	54.4-56.7	●
AM	<i>rrs</i>	a1401g	600	5	74.2	71.0-77.2	99.6	99.1-99.9	89.8	88.5-91.1	●
S	<i>rpsL</i>	K43R	358	6	62.9	58.8-66.9	98.1	95.8-99.3	75.3	72.3-78.1	●
...

















Advantages of the Grading System

- Specific mutations showed different confidence levels depending upon the medium considered for phenotypic DST
- The number of these discrepancies varied from just one variant for drugs such as AM, S, and ETO/PTO to 76 for Z

Genetic variants:



Advantages of the Grading System

Drug	Gene	Mutation	Confidence value	
			Liquid DST	Solid DST
CM	<i>rs</i>	a1401g		
R	<i>rpoB</i>	H526C		
KM	<i>eis</i>	g-10a		
H	<i>inhA</i>	c-15t		
R	<i>rpoB</i>	Q513K		
R	<i>rpoB</i>	L533P	Indeter	
R	<i>rpoB</i>	D516Y	Indeter	
MPX	<i>gyrA</i>	A90V		
R	<i>rpoB</i>	L511P		

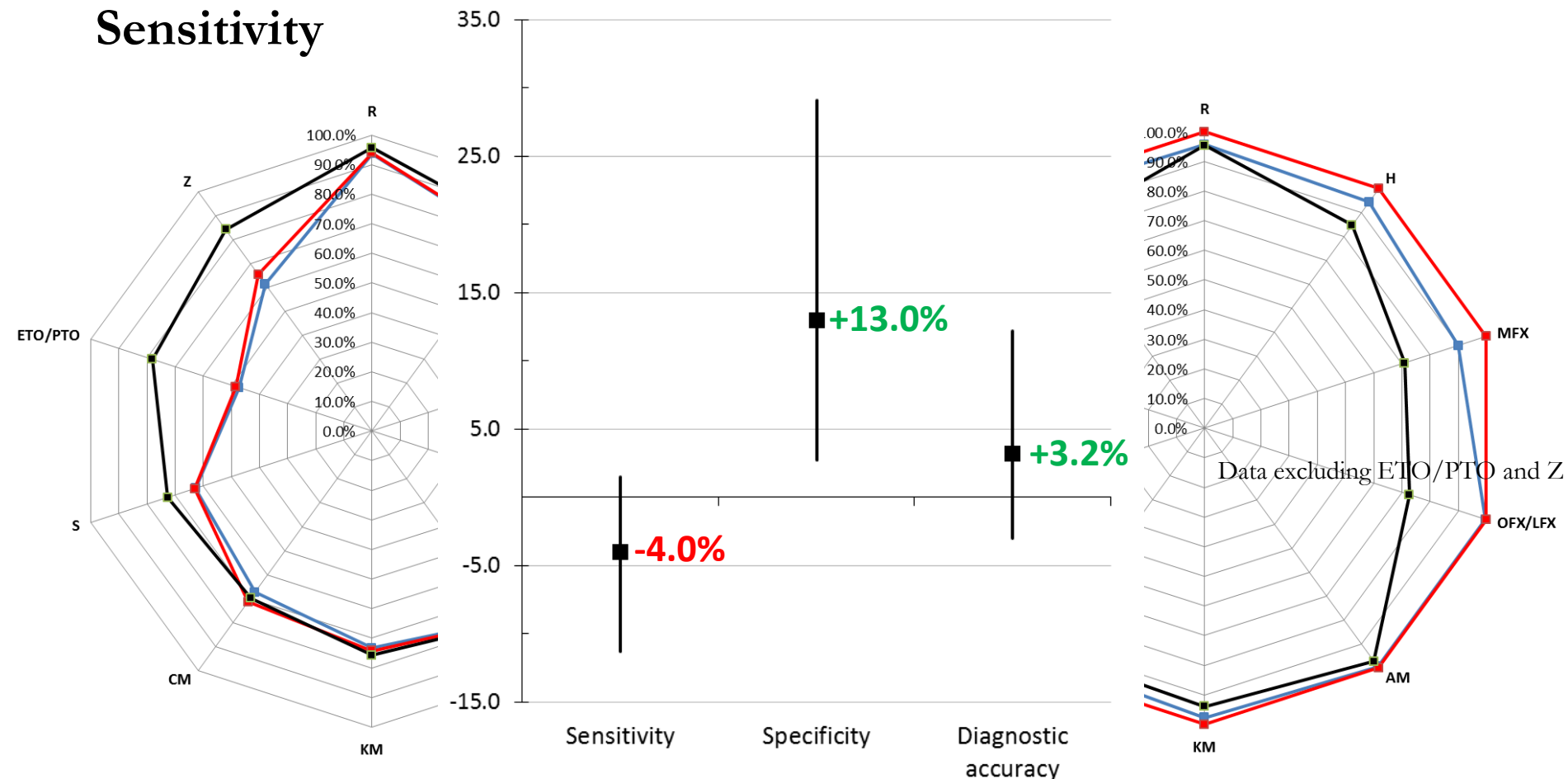
Cases testing differently
on different media

MIC close to liquid testing
critical concentration

Cases missed by liquid
phenotypic testing

Implications of the Grading System

Sensitivity



- The use of graded mutations allows for improvement in specificity with a relatively small decrease in sensitivity
- Reduction of Very Major Errors (DR reported as DS)

- A total of 389 high, moderate, and minimal confidence mutations have been determined
- Advantages of the grading system:
 - suitable for automated classification of mutations
 - LR is a universal measure of association which is not affected by local prevalence
 - unlike sensitivities and specificities, LRs do not lead to an exaggeration of the benefits of a test or strength of an association
- The validation process is driven by statistics based on frequency of mutations, this will be improved by integrating
 - Homoplasmy data
 - MIC data
 - functional genetic studies
- The validation process will need to be evaluated periodically and refined as tools and our understanding increase

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